

Determination of the Geographical Origins of Olive Mill Wastewater: FTMIR Spectroscopy and Chemometric Algorithms for Accurate Classification

Façal Sbai El Otmani[†]; Aimen El Orche^{†*}; Mouad Mouhsin[†]; Meriem Kasbaji[†]; Ayoub Ait Oumghar[†]; Mohamed Mbarki[†] and Mustapha Oubenali^{†*}

[†] Team of Analytical & Computational Chemistry, Nanotechnology and Environment, Faculty of Science and Technologies, Sultan Moulay Slimane University, BP 523, Beni Mellal, Morocco;

*e-mail: mustapha.oubenali@gmail.com

[‡] Laboratory of drug sciences, biomedical research, and biotechnology, Faculty of Medicine and Pharmacy, Hassan II University of Casablanca, Morocco.

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The present research studies the geographical classification of olive mill wastewater (OMWW) samples using FT-MIR spectroscopy and chemometric recognition algorithms. The study focuses on samples collected from two regions in Morocco: BeniMellal-Khenifra and Fes-Meknes. Principal Component Analysis (PCA) has been applied to the spectral data, revealing distinct clustering of OMWW samples based on their geographical origin. Additionally, Partial Least Squares Discriminant Analysis (PLS-DA) has been employed to develop a classification model for accurate sample categorization. The PLS-DA model presented high sensitivity and specificity, achieving perfect classification rates during the calibration phase. Validation results show the model's capability to accurately identify the majority of samples, with a minor misclassification. These results confirm the potential of FTMIR spectroscopy and chemometric recognition algorithms for authentication, and traceability in the olive oil industry. Further research can explore the extension of this methodology to other regions and agricultural by-products, incorporating advanced machine learning algorithms for enhanced classification accuracy. Overall, the present study contributes to enhance environmental waste management research by addressing OMWW disposal concerns and promoting sustainable practices on the valuation possibilities.

Keywords: Olive Mill Wastewater (OMWW), FTMIR, data analysis, traceability, classification and discrimination

Introduction

Olive oil is widely recognized as a primary fats source in the Mediterranean diet that plays a significant role in the culinary habits of Mediterranean countries. It possesses health-promoting properties thanks to its abundant content of unsaturated fatty acids. However, the production of olive oil is accompanied by a notable amount of a by-product known as olive mill wastewater (OMWW) that is generated in substantial quantities during a specific timeframe, typically spanning from October to December. Morocco, being the sixth largest producer of olive oil, generates an estimated annual volume of over 250,000 m³ of olive mill wastewater (OMWW) [1].

These OMWW possess detrimental characteristics including pollution, phytotoxicity, and antimicrobial properties, making them a significant environmental concern in the olive oil production industry [2]. Various methods have been suggested to address the challenges posed by olive mill wastewater (OMWW). These methods include evaporation ponds, thermal concentration, physico-chemical and biological

treatments, and the direct use of OMWW as an organic fertilizer in agricultural soils. Nevertheless, the two most commonly employed techniques today involve the direct application of OMWW to agricultural soils and their storage in evaporation ponds, resulting in the production of sludge [3].

For better understanding of this olive oil by-product several studies were conducted on physicochemical composition of OMWW. It is typically characterized as a dark brown to deep black discharge, exhibiting acidity and a high pollution load primarily due to substantial amounts of suspended solids and a substantial organic content. This organic content comprises polysaccharides, sugars, polyphenols, polyalcohols, tannins, proteins, organic acids and lipids [4-6]. Additionally, OMWW contains significant quantities of mineral salts, particularly potassium, phosphate and sodium [7,8]. Although limited in scope, the microbiological analysis of OMWW has revealed the presence and isolation of numerous species from yeast, fungal, and bacterial populations within these effluents [9-12]. The chemical composition of OMWW is influenced by uncontrolled multiple factors such

as olive cultivars, fruit ripening, harvesting period, storage conditions, farming methods, production origin, climatic conditions and oil extraction technology [13-18]. In fact, several studies have been carried on olive by-products as OMWW, namely in terms of biological applications of their polyphenolic compounds content [19-24].

In the present study, we develop rapid FTIR spectroscopic method combined with chemometric tools that could be able to discriminate and classify Olive Mill Wastewater samples collected from different provinces in the two Moroccan regions Beni Mellal-Khenifra and Fes-Meknes.

Material and methods

Sampling

Calibration (40) and validation (16) samples of OMWW were collected from different industrial olive oil facilities in different provinces in the two regions: Beni Mellal-Khenifra (Khouribga, Azilal, Beni Mellal and Fkih Ben Saleh) and Fes-Meknes (Taounat, Fes, Sefrou, Meknes and El Hajeb) (Table 1).

Table 1. Geographical origins of the collected samples.

Region	Province	Calibration Samples	Validation Samples
Beni Mellal - Khénifra	Khouribga	8	2
	Azilal	4	2
	Beni Mellal	7	2
	Fkih Ben Saleh	8	4
Fes - Meknes	Taounat	2	1
	Fes	3	1
	Sefrou	2	1
	Meknes	5	3
	El Hajeb	1	-
	Total	40	16

The OMWW collected were produced during the olive oil crushing period from October 2021 to January 2022 and then stored at 4 °C.

ATR-FTIR Spectroscopy

The analysis of the samples was performed by the TENSOR 27 ATR-FTIR spectrometer equipped with a horizontal ATR accessory by a Germanium crystal. Using a micropipette, each well homogenized OMWW sample was deposited on the crystalline surface of the ATR accessory. For each analysis, the ATR accessory is cleaned by an ethanol solution. The spectra were recorded in absorbance mode, collected between 4000 cm⁻¹ and 400 cm⁻¹, with 32 scans and a 4 cm⁻¹ resolution.

Multivariate Analysis

Previous studies, on olive oil produced in Morocco, have already used the coupling between spectroscopic methods, in particular FTIR, on the one hand and

methods of multivariate analysis on the other hand [25-29]. The results of these studies have motivated us to investigate into such a coupling not to olive oil as one of the products of the olive tree but rather to OMWW as one of the by-products of this plant. So, we have used various statistical techniques in the present study to process and evaluate spectral data obtained by ATR-FTIR spectroscopy. To explore and represent the data set, we first applied Principal Component Analysis (PCA). PCA allows us to search for directions of maximum dispersion in order to synthesize new variables and represent the data in a reduced-dimensional space. In addition to reducing data size, this method is frequently used to clean up data by identifying outliers. It is also an effective tool for identifying groups of individuals that behave similarly in terms of measured variables.

At the second step the principal components are used in a Partial least squares regression analysis. PLS regression is a modern method in chemometrics that enhance and couple the characteristics of principal component analysis and multiple regressions. Its objective is to predict or examine a group of dependent variables using a set of independent variables or predictors. This prediction is achieved by extracting a collection of orthogonal factors known as latent variables from the predictors that possess optimal predictive capability.

In the present study, PLS-DA was applied to find what were the variables that effectively discriminate within samples groups based on their FTIR spectra (X data block) according to their maximum covariance with a geographical location indicated by a class relevance variable (Y data block).

PLS-DA aims to determine whether a sample's spectrum corresponds to a specific geographical location, categorized as binary values of zero (0) and one (1).

The statistical processing of the data was carried out using The Unscramble Version X 10.4 software.

Results and discussion

FTIR Analysis

The FTIR absorption spectra of the collected samples are presented in Fig. 1.

The FTIR spectra show a similar appearance to that of water that is the major component of OMWW but it also contains weak absorbance's corresponding to functional groups that characterize the olive oil compounds (Table 2).

The visual observation of the ATR-FTIR spectra of these 40 samples cannot be used to determine the similarities between individuals to differentiate between OMWW coming from different geographical origins.

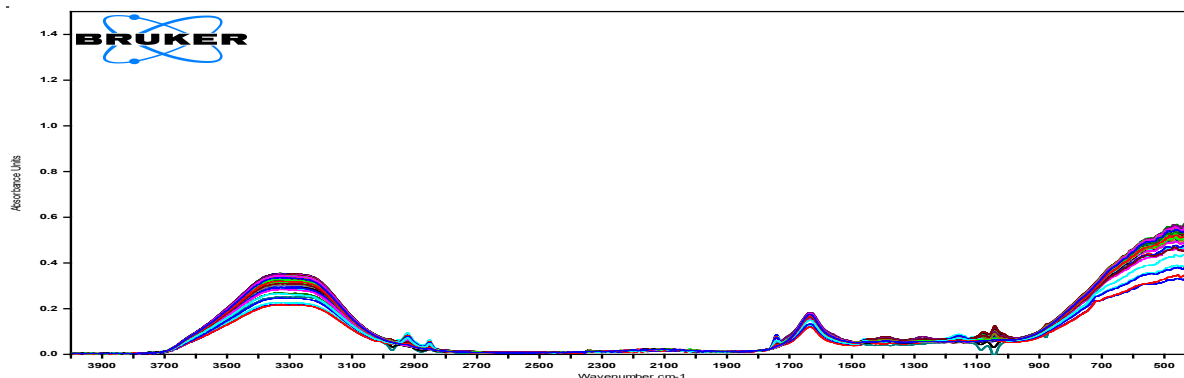


Fig. 1. ATR-FTMIR spectra of the collected OMWW samples.

Table 2. Characteristics of the ATR-FTMIR spectrum of OMWW [30, 31].

Wavenumber in cm^{-1}	Corresponding functional groups
Large and intense band at 3300	Elongation of O-H (phenol)
band at 2920	Elongation of C-H (saturated and aromatic)
pic at 1720	Elongation of C=O (Ketones)
band at 1630	Elongation of C=C unconjugated double bond

Principal Component Analysis

A preliminary examination of the spectral data collected was carried out by principal component analysis using the 400–4000 cm^{-1} FTMIR spectral range, in order to represent the data in a reduced dimensional space. The PCA results (Fig. 2) show that three principal components explain 97% of the total variance in the spectral data, accounting for 85, 8 and 4 % of the total variance in the raw data respectively.

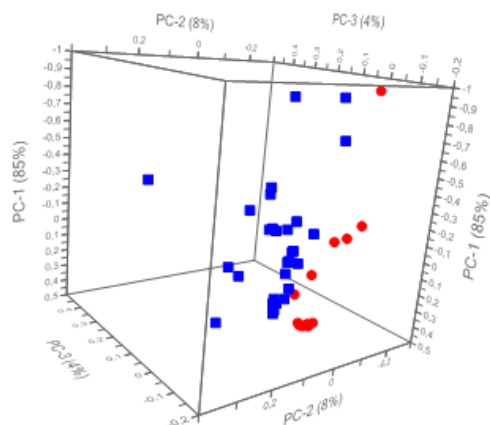


Fig. 2. 3D PCA score plot of ATR-FTMIR spectra at the PC1, PC2 and PC3 on olive mill wastewater from two different regions (Beni Mellal-Khenifra - blue, and Fes-Meknes - red).

The observation of the score plot shows that there is a clustering of OMWW based on the geographical origin of the samples. The PCA also determines that the samples clustered in two groups, the first group is representative of the Beni Mellal–Khenifra region while the second one concerns the Fes-Meknes region. We have compared the results of the present work, in terms of classification between OMWW samples from the same region, to those that have been published in the literature. So, the PCA has been used by various previous works which have been able to classify according to the geographical and varietal origins of samples of olive oil from Morocco [32,35]. Indeed, thanks to PCA El orche *et al.* were able to carry out the classification of olive oil in the Beni Mellal-Khenifra region [32] while Zeroual and *al.* have done so over the Fes-Meknes region and other regions in Morocco [33-35].

Partial Least Squares Discriminate Analysis

To develop a classification model capable of accurately categorizing samples based on their geographical origin, we employed Partial Least Squares Discriminant Analysis (PLS-DA) on spectroscopic data. PLS-DA is a robust multivariate statistical technique that combines the power of partial least squares regression with discriminant analysis. By leveraging the inherent patterns and relationships present in the spectroscopic data, PLS-DA enables us to extract meaningful features and create a predictive model. Through this approach, we aim to achieve a high level of accuracy in classifying samples based on their origin, facilitating various applications such as authentication and traceability. When analyzing the score plot (Fig. 3), we can observe distinct clusters or well-separated groups representing samples of different origins. This high level of classification in the score plot implies that the PLS-DA model has effectively learned and utilized the spectral information to differentiate samples according to their origin. It suggests that the model has extracted informative features from the spectroscopic data, allowing it to accurately classify samples based on the unique spectral signatures associated with each geographical origin.

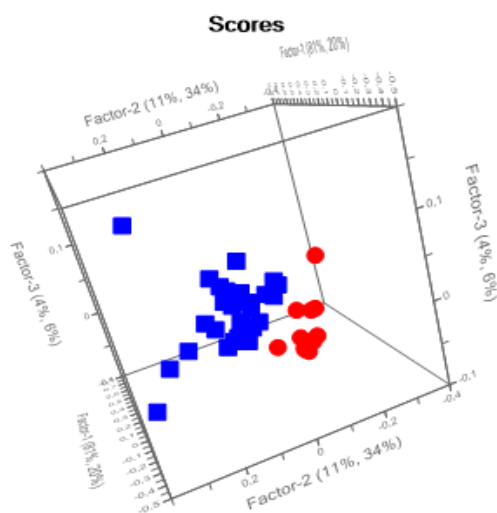


Fig. 3. PLS-DA score plot of ATR-FTMIR spectra at the PC1, PC2 and PC3 on olive mill wastewater from two different regions (Beni Mellal-Khenifra - blue, and Fes-Meknes - red).

The obtained confusion matrix from the calibration results of the PLS-DA model provides valuable insights into its performance (Table 3). The matrix reveals that all samples belonging to Class 1 (Beni Mellal-Khenifra) were accurately classified, resulting in a sensitivity of 100%. Similarly, all samples of Class 2 (Fes-Meknes) were correctly identified, leading to a specificity of 100%. These high values indicate that the model effectively distinguished between the two classes and displayed a strong capability to correctly classify samples based on their geographical origin.

Furthermore, the Correct Classification Rate (CCR) of 100% demonstrates the overall accuracy of the PLS-DA model on the given dataset. This indicates that the model achieved flawless classification, correctly assigning all samples to their respective origins. These remarkable results emphasize the robustness and reliability of the PLS-DA model when applied to spectroscopic data for classification purposes.

Analyzing the confusion matrix, we can draw valuable insights into the performance of the PLS-DA

model in classifying samples during the validation phase.

The sensitivity of 100% indicates that all samples belonging to Class 1 (Beni Mellal-Khenifra) were correctly identified by the model. This means that the model accurately detected and classified all samples originating from Class1 (Beni Mellal-Khenifra). Similarly, the specificity of 83.33% suggests that the model correctly classified a majority of the samples from Class 2 (Fes-Meknes) (Table 4). However, it did misclassify one sample from Class 2 (Fes-Meknes) as Class 1 (Beni Mellal-Khenifra). The Correct Classification Rate (CCR) measures the overall accuracy of the model and takes into account both true positives and true negatives. In this case, the CCR is calculated by summing the true positives and true negatives and dividing it by the total number of samples. Therefore, the CCR is approximately 93.75%. This indicates that the model achieved a high overall accuracy in classifying the samples during the validation phase.

These results demonstrate the effectiveness of the PLS-DA model in correctly identifying the majority of samples from both Class 1 (Beni Mellal-Khenifra) and Class 2 (Fes-Meknes). While the specificity value indicates a small misclassification, the model exhibited a strong sensitivity and a high overall accuracy (CCR) in classifying the samples based on their geographical origin. Thus the results of the present work, in terms of discrimination between OMWW samples from the same region, are comparable to those of previous work on virgin olive oil in France [36,37] and olive products in Morocco [38-41]. PLS-DA model showed a high sensitivity, indicating accurate identification of Class 1 (BeniMellal- Khenifra) samples. The specificity value suggests a mostly correct classification of Class 2 (Fes-Meknes) samples, with a minor misclassification. The CCR value reflects the overall accuracy of the model in classifying the samples during the validation phase. These results emphasize the model's capability to effectively classify samples based on their geographical origin that can have significant applications including authentication, and traceability.

Table 3. Confusion matrix of calibration results obtained by PLS-DA for the dataset.

Origin	1	2	Specificity (%)	Sensitivity (%)	CCR (%)
1	27	0	100	100	100
2	0	13			

Table 4. Confusion matrix of validation results obtained by PLS-DA for the dataset.

Origin	1	2	Specificity (%)	Sensitivity (%)	CCR (%)
1	10	1	83.33	100	93.75
2	0	5			

Conclusion

The present study has shown the successful application of FTMIR spectroscopy coupled to chemometric recognition algorithms, namely PCA and PLS-DA, to perform a geographical classification of olive mill wastewater (OMWW) samples from two geographical areas. The PCA analysis revealed distinct clustering of OMWW samples based on their geographical origin, while the PLS-DA model achieved high accuracy in accurately classifying samples according to their origin. These results highlight the potential of this approach for authentication, and traceability purposes in industries related to olive oil production. By effectively utilizing spectral data and extracting meaningful features, this methodology offers valuable insights into the classification and traceability of OMWW samples, addressing environmental concerns and supporting sustainable practices in the olive oil industry. Further research could explore the expansion of this methodology to a broader range of geographical regions and investigate its applicability to other by-products in the agricultural and food processing sectors. Additionally, integrating complementary analytical techniques and exploring advanced machine learning algorithms could further enhance the accuracy and efficiency of classification models. These advancements would contribute to improved environmental management, ensuring proper disposal of OMWW, and supporting sustainable practices in the olive oil industry and even other products or by-products.

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Conflicts of Interest

All authors declare no conflict of interest.

References

1. Hanafi, F.; Assobhei, O.; Mountadar, M. Detoxification and Discoloration of Moroccan Olive Mill Wastewater by Electrocoagulation. *J. Hazard. Mater.* **2010**, *174*(1–3), 807–812. doi:10.1016/j.jhazmat.2009.09.124
2. Paredes, C.; Cegarra, J.; Roig, A.; Sánchez-Monedero, M. A.; Bernal, M. P. Characterization of Olive Mill Wastewater (Alpechin) and Its Sludge for Agricultural Purposes. *Bioresour. Technol.* **1999**, *67*(2), 111–115. doi:10.1016/S0960-8524(98)00106-0
3. Fiestas Ros de Ursinos, J. A.; Borja Padilla, R. Use and Treatment of Olive Mill Wastewater: Current Situation and Prospects in Spain. *Grasas Aceites* **1992**, *43*(2), 101–106. doi:10.3989/gya.1992.v43.i2.1182
4. Amaral, C.; Lucas, M. S.; Coutinho, J.; Crespi, A. L.; do Rosário Anjos, M.; Pais, C. Microbiological and Physicochemical Characterization of Olive Mill Wastewaters from a Continuous Olive Mill in Northeastern Portugal. *Bioresour. Technol.* **2008**, *99*(15), 7215–7223. doi:10.1016/j.biortech.2007.12.058
5. Bouknana, D.; Hammouti, B.; Salghi, R.; et al. Physicochemical Characterization of Olive Oil Mill Wastewaters in the Eastern Region of Morocco. *J. Mater. Environ. Sci.* **2014**, *5*(4), 1039–1058.
6. Mseddi, S.; Chaari, L.; Belaid, C.; Chakchouk, I.; Kallel, M. Valorization of Treated Olive Mill Wastewater in Fertigation Practice. *Environ. Sci. Pollut. Res.* **2016**, *23*(16), 15792–15800. doi:10.1007/s11356-015-4353-6
7. Massadeh, M. I.; Modallal, N. Ethanol Production from Olive Mill Wastewater (OMW) Pretreated with *Pleurotus sajor-caju*. *Energy Fuels* **2008**, *22*(1), 150–154. doi:10.1021/ef7004145
8. Ntougias, S.; Gaitis, F.; Katsaris, P.; Skoulika, S.; Iliopoulos, N.; Zervakis, G. I. Effects of Olive Harvest Period and Production Year on Olive Mill Wastewater Properties: Evaluation of *Pleurotus* Strains as Bioindicators of Effluent Toxicity. *Chemosphere* **2013**, *92*(4), 399–405. doi:10.1016/j.chemosphere.2013.01.033
9. Sinigaglia, M.; Di Benedetto, N.; Bevilacqua, A.; Corbo, M. R.; Capece, A.; Romano, P. Yeasts Isolated from Olive Mill Wastewaters from Southern Italy: Technological Characterization and Potential Use for Phenol Removal. *Appl. Microbiol. Biotechnol.* **2010**, *87*(6), 2345–2354. doi:10.1007/s00253-010-2684-2
10. Bleve, G.; Lezzi, C.; Chiriatti, M. A.; et al. Selection of Non-Conventional Yeasts and Their Use in Immobilized Form for the Bioremediation of Olive Oil Mill Wastewaters. *Bioresour. Technol.* **2011**, *102*(2), 982–989. doi:10.1016/j.biortech.2010.09.059
11. Tsiamis, G.; Tzagkaraki, G.; Chamalaki, A.; et al. Olive-Mill Wastewater Bacterial Communities Display a Cultivar-Specific Profile. *Curr. Microbiol.* **2012**, *64*(2), 197–203. doi:10.1007/s00284-011-0049-4
12. Martínez-Gallardo, M. R.; López, M. J.; López-González, J. A.; et al. Microbial Communities of the Olive Mill Wastewater Sludge Stored in Evaporation Ponds: A Resource for Sustainable Bioremediation. *J. Environ. Manage.* **2021**, *279* (Dec 2020). doi:10.1016/j.jenvman.2020.111810
13. Casa, R.; D'Annibale, A.; Pieruccetti, F.; Stazi, S. R.; Sermanni, G. G.; Lo Cascio, B. Reduction of Phenolic Components in Olive-Mill Wastewater by Enzymatic Treatment and Its Impact on Durum Wheat (*Triticum durum* Desf.) Germinability. *Chemosphere* **2003**, *50*(8), 959–966. doi:10.1016/S0045-6535(02)00707-5
14. Aktas, E. S.; Imre, S.; Ersoy, L. Characterization and Lime Treatment of Olive Mill Wastewater. *Water*

Res. **2001**, 35(9), 2336–2340. doi:10.1016/S0043-1354(00)00490-5

15. Fiorentino, A.; Gentili, A.; Isidori, M.; et al. Environmental Effects Caused by Olive Mill Wastewaters: Toxicity Comparison of Low-Molecular-Weight Phenol Components. *J. Agric. Food Chem.* **2003**, 51(4), 1005–1009. doi:10.1021/jf020887d

16. Ben Sassi, A.; Boularbah, A.; Jaouad, A.; Walker, G.; Boussaid, A. Comparison of Olive Oil Mill Wastewaters (OMW) from Three Different Processes in Morocco. *Process Biochem.* **2006**, 41(1), 74–78. doi:10.1016/j.procbio.2005.03.074

17. El Yamani, M.; Ghabbour, N.; Sakar, E. H.; Rharrabti, Y. Olive Oil Wastewaters from Northern Morocco: Physicochemical Characterization and Antibacterial Activity of Polyphenols. *J. Mater. Environ. Sci.* **2017**, 8(8), 2667–2378.

18. El Yamani, M.; Sakar, E. H.; Boussakouran, A.; Ghabbour, N.; Rharrabti, Y. Physicochemical and Microbiological Characterization of Olive Mill Wastewater (OMW) from Different Regions of Northern Morocco. *Environ. Technol.* **2020**, 41(23), 3081–3093. doi:10.1080/09593330.2019.1597926

19. Leouifoudi, I.; Ziad, A.; Amechrouq, A.; Oukerrou, M. A.; Mouse, H. A.; Mbarki, M. Identification and Characterization of Phenolic Compounds Extracted from Moroccan Olive Mill Wastewater. *Food Sci. Technol.* **2014**, 34(2), 249–257. doi:10.1590/fst.2014.0051

20. Leouifoudi, I.; Harnafi, H.; Ziad, A. Olive Mill Waste Extracts: Polyphenols Content, Antioxidant, and Antimicrobial Activities. *Adv. Pharmacol. Sci.* **2015**, 2015, Article ID 714138. doi:10.1155/2015/714138

21. Leouifoudi, I.; Mbarki, M.; Tilaoui, M.; et al. Study of the *In Vitro* Anticancer Activity of Moroccan Phenolic Olive Cake Extracts. *J. Pharmacogn. Phytochem.* **2014**, 2(6), 154–165.

22. Elbir, M.; Es-Safi, N. E.; Amhoud, A.; Mbarki, M. Characterization of Phenolic Compounds in Olive Stones of Three Moroccan Varieties. *Maderas Cienc. Tecnol.* **2015**, 17(3), 479–492. doi:10.4067/S0718-221X2015005000043

23. El Orche, A.; Bouatia, M.; Yanisse, S.; Labjar, H.; Mouhsin, M.; Bouha, M.; Oubenali, M.; Mbarki, M. Evaluation of the Capability of Horizontal ATR-FTMIR and UV-Visible Spectroscopy in the Discrimination of Virgin Olive Oils from the Moroccan Region of Beni Mellal-Khenifra. *J. Spectrosc.* **2020**, Article ID 9317350, 1–9. doi:10.1155/2020/9317350

24. Olmo-García, L.; Bajoub, A.; Benlamaalam, S.; et al. Establishing the Phenolic Composition of *Olea europaea* L. Leaves from Cultivars Grown in Morocco as a Crucial Step Towards Their Subsequent Exploitation. *Molecules* **2018**, 23(10), 1–18. doi:10.3390/molecules23102524

25. El Orche, A.; Bouatia, M.; Mbarki, M. Rapid Analytical Method to Characterize the Freshness of Olive Oils Using Fluorescence Spectroscopy and Chemometric Algorithms. *J. Anal. Methods Chem.* **2020**,

2020, Article ID 8860161. doi:10.1155/2020/8860161

26. El Orche, A.; Bouatia, M.; Yanisse, S.; et al. Evaluation of the Capability of Horizontal ATR-FTMIR and UV-Visible Spectroscopy in the Discrimination of Virgin Olive Oils from the Moroccan Region of Beni Mellal-Khenifra. *J. Spectrosc.* **2020**, Article ID 9317350. doi:10.1155/2020/9317350

27. Laouni, A.; El Orche, A.; Elhamdaoui, O.; Karrouchi, K.; El Karbane, M.; Bouatia, M. Preliminary Study on the Potential of FT-IR Spectroscopy and Chemometrics for Tracing the Geographical Origin of Moroccan Virgin Olive Oils. *J. AOAC Int.* **2023**, 106(3), 804–812. doi:10.1093/jaoacint/qsac136

28. Hirri, A.; Bassbasi, M.; Platikanov, S.; Tauler, R.; Oussama, A. FTIR Spectroscopy and PLS-DA Classification and Prediction of Four Commercial Grade Virgin Olive Oils from Morocco. *Food Anal. Methods* **2016**, 9(4), 974–981. doi:10.1007/s12161-015-0255-y

29. Benlamaalam, S.; Chigr, M.; Hirri, A.; Auhmani, A.; Mbarki, M. Characterization and Discrimination of Virgin Olive Oils from Different Moroccan Geographical Areas Using MIR Spectroscopy Coupled to Chemometric Methods. *Int. J. Sci. Res.* **2015**, 4(6), 2443–2448. doi:10.13140/RG.2.1.4760.9443

30. Pretsch, E.; Bühlmann, P.; Affolter, C. *Structure Determination of Organic Compounds: Tables of Spectral Data*; 3rd ed.; Springer-Verlag: Berlin, Heidelberg, 2000. doi:10.1007/978-3-662-04201-4

31. Larkin, P. J. *Infrared and Raman Spectroscopy: Principles and Spectral Interpretation*; 2011. doi:10.1016/C2015-0-00806-1

32. El Orche, A.; Bouatia, M.; Labjar, H.; Maaouni, M.; Mbarki, M. Coupling Mid Infrared Spectroscopy to Mathematical and Statistical Tools for Automatic Classification, Qualification, and Quantification of Argan Oil Adulteration. *6th Int. Conf. Optim. Appl. ICOA* **2020**, Proc. 1–4. doi:10.1109/ICOA49421.2020.9094456

33. Zaroual, H.; Chénè, C.; El Hadrami, E. M.; Karoui, R. Application of New Emerging Techniques in Combination with Classical Methods for the Determination of the Quality and Authenticity of Olive Oil: A Review. *Crit. Rev. Food Sci. Nutr.* **2022**, 62(16), 4526–4549. doi:10.1080/10408398.2021.1876624

34. Zaroual, H.; Chénè, C.; Mestafa El Hadrami, E.; Karoui, R. Comparison of Four Classification Statistical Methods for Characterising Virgin Olive Oil Quality During Storage up to 18 Months. *Food Chem.* **2022**, 370 (Aug 2021). doi:10.1016/j.foodchem.2021.131009

35. Zaroual, H.; El Hadrami, E. M.; Karoui, R. Preliminary Study on the Potential Application of Fourier-Transform Mid-Infrared for the Evaluation of Overall Quality and Authenticity of Moroccan Virgin Olive Oil. *J. Sci. Food Agric.* **2021**, 101(7), 2901–2911. doi:10.1002/jsfa.10922

36. Dupuy, N.; Galtier, O.; Ollivier, D.; Vanloot,

P.; Artaud, J. Comparison between NIR, MIR, Concatenated NIR and MIR Analysis and Hierarchical PLS Model. Application to Virgin Olive Oil Analysis. *Anal. Chim. Acta* **2010**, *666*(1–2), 23–31. doi:10.1016/j.aca.2010.03.034

37. Laroussi-Mezghani, S.; Vanloot, P.; Molinet, J.; et al. Authentication of Tunisian Virgin Olive Oils by Chemometric Analysis of Fatty Acid Compositions and NIR Spectra: Comparison with Maghrebien and French Virgin Olive Oils. *Food Chem.* **2015**, *173*, 122–132. doi:10.1016/j.foodchem.2014.10.002

38. De Luca, M.; Terouzi, W.; Ioele, G.; et al. Derivative FTIR Spectroscopy for Cluster Analysis and Classification of Moroccan Olive Oils. *Food Chem.* **2011**, *124*(3), 1113–1118. doi:10.1016/j.foodchem.2010.07.010

39. De Luca, M.; Terouzi, W.; Kzaiber, F.; Ioele, G.; Oussama, A.; Ragno, G. Classification of Moroccan Olive Cultivars by Linear Discriminant Analysis Applied to ATR-FTIR Spectra of Endocarps. *Int. J. Food Sci. Technol.* **2012**, *47*(6), 1286–1292. doi:10.1111/j.1365-2621.2012.02972.x

40. El Mouftari, M.; Mahjoubi, F. Z.; Kzaiber, F.; et al. Study of Oleaster Oil's Falsification by ATR-FTIR and Chemometrics Tools. *Egypt. J. Chem.* **2021**, *64*(6), 2747–2755. doi:10.21608/ejchem.2021.53644.3107

41. Terouzi, W.; Platikanov, S.; de Juan Capdevila, A.; Oussama, A. Classification of Olives from Moroccan Regions Using Direct FT-IR Analysis: Application of Support Vector Machines (SVM). *Int. J. Innov. Appl. Stud.* **2013**, *3*(2), 493–503.